### Narrative Relation between Low Milk Somatic Cell Count and Mastitis Susceptibility in Dairy Animals

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# Abstract

Background: Somatic cell count in milk is closely associated with the incidences of intramammary infections (IMI) in milch animals, and is regarded as a reliable marker of the mammary gland (MG) infection. The contradicting school of thought exist about the interpretation and consequences of SCC; most authors reported the reduced incidences of IMI in dairy cows with lower milk SCC, but some research findings clearly explained the elevated risks linked with lower SCC, and defended in ground of compromised immune status of the focus cows. Areas Covered: The question of whether low SCC is correlated to increased susceptibility to IMI has aroused the logical and rational debate. However, to date, SCC is considered a key measure in selecting dairy animals that show reduced susceptibility to IMI. As selection indexes prioritize animals with lower SCC, it could enhance the defense mechanisms of the MG concerns that has arisen about the potential long-term increase in susceptibility to IMI. The debate surrounding this issue persists, despite seemingly contradictory findings from previous studies investigating the relationship between SCC and IMI susceptibility in dairy animals. A comprehensive understanding of the leukocyte- dynamics in milk and immune-physiology of mammary cells, along with their functions in combatting infections, can help reconcile these conflicting results. Recent studies focusing on SCCbased selection of dairy animals also contribute to resolving the matter.

Expert Opinion: It becomes apparent that the long-debated issues of susceptibility to IMI associated with SCC is more likely a phenotypic trait linked to immunomodulation rather than a consequence of selection.

Keywords: Dairy animals, Immunity, Leukocyte, Milk resident cells

# 1 Introduction

Somatic cell count (SCC) is the measure of leukocytes and alveolar epithelial casts in milk and is used as a biomarker of udder health status in milk producing animals (Matera et al., 2022; Panchal et al., 2016; Shook et al., 2017; Sun et al., 2023). Therefore, it is a proxy index for bio- surveying infected mammary gland (MG) that ensures the milk quality (Rainard et al., 2018; Schukken et al., 2003). The frequent fluctuations in SCC proportion are basically due to the rate of migration of leukocytes to the milk from the bloodstream, often in response to microbial invasion in the MG and desquamation of its epithelium (Rainard *et al.*, 2008). These leukocytes primarily function as an initial line of defense in the MG, as evidenced by development of acute intramammary infection (IMI) when the infiltration of leukocytes is impeded (Prin-Mathieu et al., 2002; Vangroenweghe et al., 2005). Moreover, the SCC, or its different form, log SCC and somatic cell score/SCS (Ali & Shook, 1980), serves as a reliable selection criterion for reducing mastitis susceptibility genetically (Odegard *et al.*, 2003). However, most of the selection based on estimated breeding values for mastitis classically rely on log-SCC, without imposing a lower limit; and uses mean lactation SCC commonly because of its moderate heritability ( 0.15) and widespread data availability (Rupp & Foucras, 2010). Having significant correlation with subclinical mastitis (SCM), use of SCC has an innate contradiction of being poorly associated with acute clinical mastitis (CM), especially those due to coliforms (Rainard *et al.*, 2018). Nonetheless, a robust genetic correlation, averaging 0.6 to 0.7, exists between SCC and CM, validating SCC's role as a surrogate for CM too (Govignon-Gion et al., 2016; Leitner et al., 2024). However, past literatures had emphasized that these two mastitis indicators represent distinct traits (Mrode and Swanson, 1996).

Consensus aligns on the selection against high SCC, yet ideas diverge regarding the consequence of low SCC (Beaudeau et al., 2002; Rainard et al., 2022a; Sharma et al., 2011). However, the issues regarding the potential risks associated with the selection of animals with extremely low SCC which may predispose them to IMIs, both SCM and CM, is a matter of debate (Beaudeau et al., 2002). Some researchers advocate for maintaining a reasonable concentration of somatic cells in milk to mitigate these risks ((Leitner et al., 2024; Schukken et al., 2003) while others argue that there is no clear lower limit to SCC that is correlated with a lower incidence of IMI (Philipsson et al., 1995; Rainard et al., 2018). Furthermore, the concept has been introduced that SCC



comprises two distinct components: the baseline SCC influenced by physiological and environmental factors, and the next linked to vulnerability to IMI (Odegård et al., 2005). Additionally, Heringstad et al. (2006) reported 0.03 and 0.08 heritability of SCS for infected and healthy dairy cows, with higher correlation (0.78) between these traits and stated that SCC may not be uniform in cows with and without mastitis. Numerous literatures have discussed the relationship between low SCC and mastitis susceptibility, yielding seemingly contradictory results (Rainard et al., 2018; Van den Borne et al., 2011). In spite of these analyses, the question remains unresolved to clarify either lower SCC predisposes the producing dairy animals to IMI or not. To navigate this apparent deadlock, it is crucial to consider other factors and explore the possible mechanisms that immune cells and MG tissue employ to fight infections. By doing so, it becomes apparent that the seemingly conflicting previous works are not necessarily irreconcilable. Furthermore, recent researches on dairy cows selected based on SCS contribute new insights to key aspects of the issue, offering a basis for a compelling resolution (Ezzat Alnakip et al., 2014)



Figure 1: Association of major determinants of the udder health [source: Rainard et al. (2018)].

#### 1.1 Variation on concept, thought and scientific reasoning

Conflicting findings from previous research works have explored the link between SCC levels and mastitis resistance. The challenge in obtaining a clear picture arises from the diverse study methodologies, which involve recording CM or SCM, bulk tank SCC, udder SCC, or teat SCC, often without detailed bacteriological analyses (Rainard et al., 2018). CM is commonly diagnosed and monitored, whereas SCM is estimated through SCC across different levels of lactation and parity (Alhussien et al., 2021; Alhussien & Dang, 2020). The multifaceted nature of various independent and interconnected affecters ultimately determines the UH status (Figure 1). In spite of this complexity, valuable information can be obtained from a diverse collection of statistics and ideas.

The association of incidences of SCC and CM appears to be influenced by the type of infection in the herd (Shook et al., 2017). For instance, Erskine et al. (1988) reported a higher incidence of CM in the high SCC cows when comparing the risks of occurrence of IMI in low ( $< 150,000 cells/mL$ ) and high ( $> 750,000 cells/mL$ ) bulk milk SCC producing herds. The high SCC cows experienced higher incidence of CM and showed a strong affinity with streptococci and staphylococci while coliforms were dominant in low SCC herds (Rainard et al., 2018). However, other research results yielded inconsistent results; some of them had reported no association between bulk milk SCC and CM, while others reported a higher incidence in herds with low SCC (Alhussien et al., 2021; Beaudeau et al., 2002; Li et al., 2014). The variation in CM incidence in herds with low SCC may be considerable, and the link between CM and low bulk milk SCC is not well established.

At the cow level, the incidence of CM was observed to increase as SCC decreased in some studies, indicating a higher risk in cows or herds having low SCC, i.e.  $< 50-200\times10^{3}$  cells/mL (Beaudeau et al., 2002; Suriyasathaporn et al., 2000). Tadich et al. (1998) and Barkema et al. (1998) too had suggested that CM tends to be more severe in herds or in cows with low SCC. While various factors, such as a higher incidence of Gm-ve bacterial infections, may contribute to these findings, but may partly be attributed to a reduced immunity of cows to manage infections when their SCC is low. However, a reduced risk of CM in a low SCC cow were reported in contradictory past studies. Steeneveld et al. (2008) reported a lower prevalence of CM in low bulk milk SCC herds than in moderately high bulk milk SCC ones, while others reported that cows were at the higher risk of IMI when they have very low SCC in milk. The variations in the study design, population characteristics, and specific SCC thresholds considered may be the cause for these conflicting findings (Petzer *et al.*, 2017; Sumon *et* al., 2020). But these incompatible and the challenging literatures offer a solid basis in synthesizing the valuable insights into the relationship between SCC levels and mastitis incidence.



# 2 Milk Somatic Cells as Immune Tool

The MG leukocytes offer a level of cellular defense against microbial invasion by triggering an instant inflammatory response to the IMI (Ezzat Alnakip et al., 2014). In addition, it secretes different immune-mediators like cytokines, chemokines, reactive oxygen species (ROS), and antimicrobial proteins in milk and helps in regulation of the inflammatory processes (Singh et al., 2008). Leukocytes form the majority of somatic cells with a small fractions of epithelial casts from the MG (Concha, 1986). Therefore, lymphocytes, macrophages, and a limited proportion of neutrophil granulocytes are the primary somatic cells in healthy glands (Dosogne *et al.*, 2003). They work as a surveillance line against infective agents and assist in apoptosis of the MG (Barber *et al.*, 1999). Specifically, neutrophils play a crucial role in controlling bacterial proliferation within the MG lumen (Paape et al., 2002) and there is a prominent shift from mononuclear to polymorphonuclear cells in the somatic cell picture during IMI (Table 1) (Sarikaya et al., 2006). As a result, selection against high SCC is essentially a selection against inflammation and neutrophil concentration in milk, while selection against low SCC implies a choice against the concentration of macrophages and lymphocytes. This perspective has led some researchers to propose that high and low SCC could be considered as distinct traits (Madsen et al., 2008). Concerns have been raised about the wisdom of selecting for very low SCC, as it might compromise the mammary gland's ability to combat infection (Kehrli & Shuster, 1994). Therefore, it is important to understand dynamics of milk leukocytes by distinguishing the roles of macrophages and lymphocytes from that of neutrophils, as well as considering the involvement of milk resident cells to evaluate the validity of this concern.

#### 2.1 Role and behavior of Milk Resident Cells

Milk resident cells are immune cells in milk of healthy MGs (Dosogne *et al.*, 2001), and are regularly refilled in milk on each milking, remaining for a short period within the MG lumen (Rainard *et al.*, 2018). More specifically, these cells are evident in the gland cisterns and ducts of the MG where milk flow flushes them out continuously during milking. The composition of the resident cell analyzed in milk evident the continual recruitment of leukocytes from the blood and MG tissue into the lumen ( $Ezzat Alnakip et al., 2014$ ). It is crucial to recognize that this resident cell mass may not precisely represent the leukocyte residing within the mammary tissue. Despite this, these migrating cells are considered the first line of defense on detecting invading alien agent, operating phagocytosis, and initiating inflammatory processes (Ezzat Alnakip *et al.*, 2014). The concentration of these cells is an important factor in their defensive role, as they rely on encounters facilitated by concentration and convection currents, rather than active movement toward targets (Lengi et al., 2022). The frequency of contact with bacteria and other microbes determines cell activation, chemotaxis, phagocytosis efficiency and dynamics of IMI in the MG.

In-vitro experiments have established that a threshold concentration of phagocytes, approximately  $5 \times 10^5$  cells/mL, is necessary to achieve a significant reduction in bacterial numbers (Li et  $al., 2002$ ). Given that MG infections are initially detected in cisterns and large ducts (Petzl  $et al., 2016$ ), these cells become particularly valuable at these sites. However, examinations of the epithelium lining the cisterns and large ducts through histological and immunohistochemical methods have not revealed macrophage-like cells adhering to the epithelium. Therefore, the existence of a substantial population of adhered macrophages in these areas remains to be confirmed (Maxymiv et al., 2012; Rainard et al., 2013).

The bacterial products (metabolites, exotoxins, exosomes, etc.) in milk too can activate milk leukocytes, during the weaker trigger of immune response to infection. Then the activity of these cells is an important point to consider. Milk macrophages have significantly lower phagocytic capacity than blood monocytes or neutrophils (Denis et al., 2006; Dosogne et al., 2001) and are less responsive to IFN- $\gamma$  and produce less IL-1 $\beta$ , TNF- $\alpha$ , as well as nitric oxide compared to blood monocytes (Denis et al., 2006; Politis et al., 1992). The functions of milk lymphocytes are not well defined, with most displaying the morphometry of T cells, mainly CD8+ T cells (Soltys & Quinn, 1999). Overall, the interaction of lymphocytes with other effector cells relies on random encounter and depend on its concentration in milk. Thus, the lower number of leukocytes in milk in a healthy MG are unlikely to play a significant role in defence.

#### 2.2 Neutrophils and the concept of the leukocyte barrier

At the initiation of inflammation in the MG, there is a profound alteration in the cellular composition of milk (Rainard et al., 2022a; Rainard et al., 2022b). Notably, the MG demonstrates a remarkable ability to rapidly initiate extensive inflammatory responses dominated by neutrophils. This results in a significant increase of neutrophils in the milk, with concentrations often exceeding  $100 \times 10^8$ cells/mL (Schalm & Lasmanis, 1968). Even in cases of persistent Staphylococcus aureus-induced chronic mastitis, neutrophils remain predominant in the milk (Riollet et al., 2001). Neutrophils play an important role in controlling IMI caused by most mastitis pathogens (Paape et al., 2002). As mentioned earlier, maintaining an optimum level of neutrophils is essential for reduction and elimination of bacteria in fluid medium, and this concentration (approximately  $3-5\times10^5$ cells/mL) is readily achieved and sustained in the inflamed MG. Schalm et al. (1967) suggested that pre-existing leukocytosis



in milk could hinder the establishment of infection following bacterial invasion into the MG lumen, proposing the concept of a leukocyte barrier to infection mediated by neutrophils resulting from local inflammation (Schalm & Lasmanis, 1968). According to the leukocyte barrier concept, a healthy MG is more susceptible to new IMI compared to an already infected and inflamed gland. A logical implication of the leukocyte barrier effect is that herds with low levels of IMI, when exposed to the same infectious pressure, are inherently more vulnerable to new infections than herds with high levels of chronic IMI (Ezzat Alnakip et al., 2014; Farschtschi et al., 2022; Jiang et al., 2022; Lozada-Soto et al., 2020).

Table 1: The differences in distribution of cellular components in mammary gland (MG) between healthy and inflammatory conditions

	<b>Healthy MG</b>	Mastitic MG
<b>SCC</b>	Usually lower than $1 \times 10^5$ cells/mL milk. However, a SCC higher than $2 \times 10^5$ cells/ mL milk is considered to be a more practical distinguishing threshold for IMI.	SCC is greater than $2 \times 10^5$ cells/mL milk according to severity of IMI; with severe IMIs, the SCC may reach $1 \times 10^6$ cells/mL references milk or more within a few hours.
Leukocytes	75% of SCC.	Dramatic increase occurs according to severity of IMI at early stages due to recruitment of immune cells from the marginal pool and bone marrow into the MG environment.
Macrophages	35-79% of total leukocytes in milk, constituting the predominant cell type.	$9-32\%$ of total leukocytes in milk.
Lymphocytes	$10-28\%$ of total leukocytes in milk. The proportions of T- and B-lymphocytes in milk are approximately 40–50% and $20-25\%$ , respectively.	$14-24\%$ of total leukocytes in milk. $CD4+$ T-cells become the predominant activated phenotype in response to recognition of Ag-MHC class II complexes on Ag-presenting cells, such as B-cells or macrophages.
	$\alpha\beta$ T-cells prevail and are predominantly $CD8+$ subset with memory characteristics (comprising approximately 50–60%) of the T-lymphocyte population).	In some circumstances, such as chronic Staph. aureus IMIs, CD8+ are predominantly recruited compared over CD4+ T-lymphocytes.
<b>PMNs</b>	$3-26\%$ of total leukocytes in milk.	The predominant cell type, constituting up to 90% of the total milk leukocytes or more. With chronic bacterial IMIs, PMNs also remain as the predominant cells, even for months.

Source: Ezzat Alnakip et al. (2014)

# 3 The Mammary Epithelium Defense

Numerous studies have confirmed the competence of mammary epithelial cells (MEC) to detect the microorganism and its products (Lahouassa et al., 2007; Rainard et al., 2022a; Yang et al., 2008). The various recognition receptors present in MEC is its identity that read different microbial pattern called microbe-associated molecular patterns (MAMPs) (Jungi et al., 2011). Transmembrane proteins, nucleotide-binding oligomerization domain (NOD)-like receptors (NOD1 and NOD2) as well as toll-like receptors are the major pattern recognition receptors (Porcherie et al., 2012). NOD1 is sensitive to a dipeptide (iE-DAP) of peptidoglycan layer in Gm-ve bacteria, while NOD2 responds to muramyl dipeptide of all peptidoglycans (Rainard *et al.*, 2022a). After detecting these MAMPs, different immune mediators such as antimicrobial peptides, reactive oxygen species, cytokines and chemokines are secreted as a MEC response (Bougarn et al., 2011).

In addition to epithelial cells, the mammary epithelium lining the cisterns and large ducts includes dendritic cells, macrophages, and lymphocytes that closely associated with the epithelial linings (Maxymiv et al., 2012). The cooperation between MEC and these epithelium-associated leukocytes is likely to amplify the response of MEC to bacteria (Bougarn et al., 2010). Notably, the reaction of MEC to Escherichia coli and S. aureus aligns well with the initial response of the MG, surpassing the response of macrophages to Streptococcus uberis



(Gilbert et al., 2013; Günther et al., 2016). This supports the notion that milk leukocytes are not the initiators of MG inflammation; instead, MEC stimulation is essential to initiate the mammary tissue response. The collaboration between MEC and tissue leukocytes is particularly effective near the entry points of pathogens, such as Furstenberg's rosette at the distal end of the teat and the teat sinus. In response to infection, these sentinels of the MG produce chemokines, cytokines, and antimicrobial proteins (Lind *et al.*, 2015; Rinaldi *et al.*, 2010) and combat the existing infection and reduces the risks of evading new ones.

In spite of the prominent role of the mammary epithelium and the presumed low contribution of milk leukocytes to inflammation initiation, observations suggest that low SCC mammary glands may not be less susceptible to IMI than high SCC glands (Jiang et al., 2022; Rainard et al., 2018) and this contradicts common observations. The SCC could potentially serve as an indicator of the reactivity of the MG to bacterial stimuli, not because milk cells trigger the reaction, but due to higher concentrations of migrated cells reflecting higher concentrations of tissue-associated cells. Moreover, beyond a critical concentration, inflammatory milk cells (distinct from "resident cells" in uninflamed glands) might contribute to inflammation initiation (Fromageau et al., 2011; Rainard et al., 2022a; Rainard et al., 2022b).



Figure 2: The mammary gland epithelium at the level of cisterns and large ducts. MG epithelium can recognize bacteria and initiate the inflammatory responses. Dendritic cells/macrophages and cluster of differentiation (CD) 8+ T cell is closely associated with the epithelium, adjacent to mammary epithelial cells (MEC) and others in the sub-epithelial tissue. Source: (Rainard et al., 2018)

### 4 Resistance to Mastitis and Low Scc

Literature exploring the correlation between low SCC and susceptibility to mastitis presents divergent findings (Beaudeau et al., 2002; Rainard et al., 2018). The apparent contradictions in the interpretation of study results can be attributed to the involvement of various mechanisms of underlying IMI:

- The immune barrier formed by milk leukocytosis linked to pre-existing infections
- Infection pressure arising from exposure to diverse pathogens, be they of contagious or environmental origin
- The level of genetic susceptibility of the host to MG infections
- MG reactivity influenced by current or past environmental factors

The dichotomy between resistance and susceptibility to infection is a transient phenotypic trait of the udder, primarily dictated by the physiological state of the animal and its genetic makeup. A crucial factor influencing the susceptibility trait is the presence of prior or concurrent udder infections, inducing persistent innate immunity, termed trained immunity (Netea *et al.*, 2011), as seen with that of the leukocyte barrier. This immunity is largely individual-centric, albeit some effects extend to the entire udder (Bisutti et al., 2023; Rainard et al., 2022b; Vlasova & Saif, 2021).

A significant outcome of the local expression of resistance to infection is that herds with a higher proportion of chronic-infected glands exhibit a lower incidence of new infections. Conversely, herds with a greater percentage of healthy glands face an increased risk of new infections (Rainard *et al.*, 2018). The incidence of CM in a herd with low IMI levels, typically characterized by a low bulk milk SCC, is determined by the prevailing infection pressure. Consequently, low SCC quarters and herds might appear more susceptible to mastitis in epidemiological surveys,



not necessarily indicating an elevated genetic susceptibility in low SCC animals (Beaudeau et al., 2002; Rainard et al., 2018; Rupp et al., 2000).

The primary goal of genetic selection against mastitis is to augment the proportion of animals with low IMI levels and fewer instances of CM. This inadvertently promotes herds with a higher prevalence of phenotypically receptive glands. Consequently, the MG of a low SCC animal is more receptive phenotypically due to its noninflamed nature, making it less susceptible to mastitis owing to genetic resistance, compared to an infected high SCC cow (Koivula et al., 2005; Leitner et al., 2024).

### 5 Conclusion

Low SCC is not necessarily an indicator of high susceptibility to mastitis. Previous research reports suggested that increased SCC correlates with heightened MG reactivity due to existing basal inflammation, possibly indicating an alert state in response to local innate immunity. While this heightened reactivity could be advantageous in specific conditions, it entails inflammation and associated drawbacks. In other hand, mastitis susceptibility due to very low SCC may be a phenotypic trait related to immunomodulation, opening area for further well-designed works and not solely a result of selection. Regarding intentional selection for low SCC in dairy animals, researches with divergent selection demonstrate that it does not negatively impact susceptibility to MG infection. Therefore, we can conclude that the SCC of healthy and uninflamed glands does not differ due to SCS- based selection and does not reduce the reactivity to infection but the herds with existing basal bacterial invasions may experience increased risks of mastitis in dairy animals.

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#### Author Contributions

Conceptualization, methodology investigation writing original draft preparation writing review and editing all works done by DKC.

### References

- Alhussien, M. N., & Dang, A. K. (2020). Sensitive and rapid lateral-flow assay for early detection of subclinical mammary infection in dairy cows. Scientific Reports, 10(1). https://doi.org/10.1038/s41598-020-68174-0
- Alhussien, M. N., Panda, B. S. K., & Dang, A. K. (2021). A Comparative Study on Changes in Total and Differential Milk Cell Counts, Activity, and Expression of Milk Phagocytes of Healthy and Mastitic Indigenous Sahiwal Cows. Frontiers in Veterinary Science, 8, 1–12. https://doi.org/10.3389/fvets.2021.670811
- Ali, A. K. A., & Shook, G. E. (1980). An Optimum Transformation for Somatic Cell Concentration in Milk. Journal of Dairy Science, 63(3), 487–490. https://doi.org/10.3168/jds.S0022-0302(80)82959-6
- Barber, M. R., Pantschenko, A. G., Hinckley, L. S., & Yang, T. J. (1999). Inducible and Constitutive In Vitro Neutrophil Chemokine Expression by Mammary Epithelial and Myoepithelial Cells. In Clinical and Diagnostic Laboratory Immunology, 6(6). https://journals.asm.org/journal/cdli
- Barkema, H. W., Schukken, Y. H., Lam, T. J. G. M., Beiboer, M. L., Wilmink, H., Benedictus, G., & Brand, A. (1998). Incidence of Clinical Mastitis in Dairy Herds Grouped in Three Categories by Bulk Milk Somatic Cell Counts. Journal of Dairy Science, 81(2), 411–419. https://doi.org/10.3168/jds.S0022-0302(98)75591-2
- Beaudeau, F., Fourichon, C., Seegers, H., & Bareille, N. (2002). Risk of clinical mastitis in dairy herds with a high proportion of low individual milk somatic-cell counts. Preventive Veterinary Medicine, 53, 43–54.
- Bisutti, V., Mach, N., Giannuzzi, D., Vanzin, A., Capra, E., Negrini, R., Elena Gelain, M., Cecchinato, A., Ajmone-Marsan, P., & Pegolo, S. (2023). Transcriptome-wide mapping of milk somatic cells upon subclinical mastitis infection in dairy cattle. Journal of Animal Science and Biotechnology, 14, 93. https://doi.org/10.1186/s40104-023-00890-9ï
- Bougarn, S., Cunha, P., Gilbert, F. B., Harmache, A., Foucras, G., & Rainard, P. (2011). Staphylococcalassociated molecular patterns enhance expression of immune defense genes induced by IL-17 in mammary



epithelial cells. Cytokine, 56(3), 749–759. https://doi.org/10.1016/J.CYTO.2011.09.020

- Bougarn, S., Cunha, P., Harmache, A., Fromageau, A., Gilbert, F. B., & Rainard, P. (2010). Muramyl dipeptide synergizes with Staphylococcus aureus lipoteichoic acid to recruit neutrophils in the mammary gland and to stimulate mammary epithelial cells. Clinical and Vaccine Immunology, 17(11), 1797–1809. https://doi.org/10.1128/CVI.00268-10
- Concha, C. (1986). Cell types and their immunological functions in bovine mammary tissues and secretions–a review of the literature. Nordisk Veterinaermedicin, 38(5), 257–272.
- Denis, M., Parlane, N. A., Lacy-Hulbert, S. J., Summers, E. L., Buddle, B. M., & Wedlock, D. N. (2006). Bactericidal activity of macrophages against Streptococcus uberis is different in mammary gland secretions of lactating and drying off cows. Veterinary Immunology and Immunopathology, 114(1–2), 111–120. https://doi.org/10.1016/J.VETIMM.2006.08.001
- Dosogne, H., Vangroenweghe, F., Barrio, B., Rainard, P., & Burvenich, C. (2001). Decreased number and bactericidal activity against Staphylococcus aureus of the resident cells in milk of dairy cows during early lactation. Journal of Dairy Research, 68, 539–549. https://doi.org/10.1017§0022029901005088
- Dosogne, H., Vangroenweghe, F., Mehrzad, J., Massart-Leën, A. M., & Burvenich, C. (2003). Differential leukocyte count method for bovine low somatic cell count milk. Journal of Dairy Science, 86(3), 828–834. https://doi.org/10.3168/jds.S0022-0302(03)73665-0
- Erskine, R. J., Eberhart, R. J., Hutchinson, L. J., Spencer, S. B., & Campbell, M. A. (1988). Incidence and types of clinical mastitis in dairy herds with high and low somatic cell counts. Journal of the American Veterinary Medical Association, 192(6), 761–765.
- Ezzat Alnakip, M., Quintela-Baluja, M., Böhme, K., Fernández-No, I., Caamaño-Antelo, S., Calo-Mata, P., & Barros-Velázquez, J. (2014). The Immunology of Mammary Gland of Dairy Ruminants between Healthy and Inflammatory Conditions. Journal of Veterinary Medicine, 2014, 1–31. https://doi.org/10.1155/2014/659801
- Farschtschi, S., Mattes, M., & Pfaffl, M. W. (2022). Advantages and Challenges of Differential Immune Cell Count Determination in Blood and Milk for Monitoring the Health and Well-Being of Dairy Cows. In Veterinary Sciences, 9(6). MDPI. https://doi.org/10.3390/vetsci9060255
- Fromageau, A., Cunha, P., Gilbert, F. B., & Rainard, P. (2011). Purified Staphylococcus aureus leukotoxin Luk-M/F' does not trigger inflammation in the bovine mammary gland. Microbial Pathogenesis, 51(6), 396–401. https://doi.org/10.1016/j.micpath.2011.09.005
- Gilbert, F. B., Cunha, P., Jensen, K., Glass, E. J., Foucras, G., Robert-Granié, C., Rupp, R., & Rainard, P. (2013). Differential response of bovine mammary epithelial cells to Staphylococcus aureus or Escherichia coli agonists of the innate immune system. Veterinary Research, 44(1). https://doi.org/10.1186/1297-9716-44-40
- Govignon-Gion, A., Dassonneville, R., Baloche, G., & Ducrocq, V. (2016). Multiple trait genetic evaluation of clinical mastitis in three dairy cattle breeds. Animal, 10(4), 558–565. https://doi.org/10.1017/S1751731115002529
- Günther, J., Koy, M., Berthold, A., Schuberth, H. J., & Seyfert, H. M. (2016). Comparison of the pathogen species-specific immune response in udder derived cell types and their models. Veterinary Research, 47(1). https://doi.org/10.1186/s13567-016-0307-3
- Heringstad, B., Gianola, D., Chang, Y. M., Ødegård, J., & Klemetsdal, G. (2006). Genetic associations between clinical mastitis and somatic cell score in early first-lactation cows. Journal of Dairy Science, 89(6), 2236–2244. https://doi.org/10.3168/jds.S0022-0302(06)72295-0
- Jiang, L., Sun, H., Gu, F., He, J., Zhao, F., & Liu, J. (2022). Blood neutrophil extracellular traps: a novel target for the assessment of mammary health in transition dairy cows. Journal of Animal Science and Biotechnology, 13(1). https://doi.org/10.1186/s40104-022-00782-4
- Jungi, T. W., Farhat, K., Burgener, I. A., & Werling, D. (2011). Toll-like receptors in domestic animals. Cell and Tissue Research, 343(1), 107–120. https://doi.org/10.1007/s00441-010-1047-8



- Kehrli, M. E., & Shuster, D. E. (1994). Factors Affecting Milk Somatic Cells and Their Role in Health of the Bovine Mammary Gland. Journal of Dairy Science, 77(2), 619–627. https://doi.org/10.3168/jds.S0022- 0302(94)76992-7
- Koivula, M., Mäntysaari, E. A., Negussie, E., & Serenius, T. (2005). Genetic and phenotypic relationships among milk yield and somatic cell count before and after clinical mastitis. Journal of Dairy Science, 88(2), 827–833. https://doi.org/10.3168/jds.S0022-0302(05)72747-8
- Lahouassa, H., Moussay, E., Rainard, P., & Riollet, C. (2007). Differential cytokine and chemokine responses of bovine mammary epithelial cells to Staphylococcus aureus and Escherichia coli. Cytokine, 38(1), 12–21. https://doi.org/10.1016/J.CYTO.2007.04.006
- Leitner, G., Blum, S. E., Krifucks, O., Lavon, Y., Jacoby, S., & Seroussi, E. (2024). Alternative Traits for Genetic Evaluation of Mastitis Based on Lifetime Merit. Genes, 15(1). https://doi.org/10.3390/genes15010092
- Lengi, A. J., Stewart, J. W., Makris, M., Rhoads, M. L., & Corl, B. A. (2022). Heat Stress Increases Mammary Epithelial Cells and Reduces Viable Immune Cells in Milk of Dairy Cows. Animals, 12(20). https://doi.org/10.3390/ani12202810
- Li, N., Richoux, R., Boutinaud, M., Martin, P., & Gagnaire, V. (2014). Role of somatic cells on dairy processes and products: A review. Dairy Science and Technology, 94(6), 517–538. https://doi.org/10.1007/s13594- 014-0176-3
- Li, Y., Karlin, A., Loike, J. D., & Silverstein, S. C. (2002). A critical concentration of neutrophils is required for effective bacterial killing in suspension. Proceedings of the National Academy of Sciences of the United States of America, 99(12), 8289–8294. www.pnas.orgcgidoi10.1073pnas.122244799
- Lind, M., Sipka, A. S., Schuberth, H. J., Blutke, A., Wanke, R., Sauter-Louis, C., Duda, K. A., Holst, O., Rainard, P., Germon, P., Zerbe, H., & Petzl, W. (2015). Location-specific expression of chemokines, TNF-α and S100 proteins in a teat explant model. Innate Immunity, 21(3), 322–331. https://doi.org/10.1177/1753425914539820
- Lozada-Soto, E., Maltecca, C., Anderson, K., & Tiezzi, F. (2020). Analysis of milk leukocyte differential measures for use in management practices for decreased mastitis incidence. Journal of Dairy Science, 103(1), 572–582. https://doi.org/10.3168/jds.2019-16355
- Madsen, P., Shariati, M. M., & Ødegard, J. (2008). Genetic analysis of somatic cell score in danish holsteins using a liability-normal mixture model. Journal of Dairy Science, 91(11), 4355–4364. https://doi.org/10.3168/jds.2008- 1128
- Matera, R., Di Vuolo, G., Cotticelli, A., Salzano, A., Neglia, G., Cimmino, R., D'Angelo, D., & Biffani, S. (2022). Relationship among Milk Conductivity, Production Traits, and Somatic Cell Score in the Italian Mediterranean Buffalo. Animals, 12(17). https://doi.org/10.3390/ani12172225
- Maxymiv, N. G., Bharathan, M., & Mullarky, I. K. (2012). Bovine mammary dendritic cells: A heterogeneous population, distinct from macrophages and similar in phenotype to afferent lymph veiled cells. Comparative Immunology, Microbiology and Infectious Diseases, 35(1), 31–38. https://doi.org/10.1016/J.CIMID.2011.09.009
- Netea, M. G., Quintin, J., & Van Der Meer, J. W. M. (2011). Trained immunity: A memory for innate host defense. In Cell Host and Microbe, 9(5), 355–361. https://doi.org/10.1016/j.chom.2011.04.006
- Odegård, J. a, Klemetsdal, G., & Heringstad, B. (2003). Genetic improvement of mastitis resistance: Validation of somatic cell score and clinical mastitis as selection criteria. Journal of Dairy Science, 86(12), 4129–4136. https://doi.org/10.3168/jds.S0022-0302(03)74027-2
- Odegård, J., Madsen, P., Gianola, D., Klemetsdal, G., Jensen, J., Heringstad, B., & Korsgaard, I. R. (2005). A Bayesian threshold-normal mixture model for analysis of a continuous mastitis-related trait. Journal of Dairy Science, 88(7), 2652–2659. https://doi.org/10.3168/jds.S0022-0302(05)72942-8
- Paape, M., Mehrzad, J., Zhao, X., Detilleux, J., & Burvenich, C. (2002). Defense of the bovine mammary gland by polymorphonuclear neutrophil leukocytes. In Journal of Mammary Gland Biology and Neoplasia, 7(2), 109–121.



https://doi.org/10.1023/A:1020343717817

- Panchal, I., Sawhney, I., & Dang, A. (2016). Relation between electrical conductivity, dielectric constant, somatic cell counts and some other milk quality parameters in diagnosis of subclinical mastitis in Murrah buffaloes I Panchal, IK Sawhney and AK Dang. Indian Journal of Dairy Science, 69(3), 267–271.
- Petzer, I. M., Karzis, J., Donkin, E. F., Webb, E. C., & Etter, E. M. C. (2017). Somatic cell counts thresholds in composite and quarter milk samples as indicator of bovine intramammary infection status. Onderstepoort Journal of Veterinary Research, 84(1). https://doi.org/10.4102/ojvr.v84i1.1269
- Petzl, W., Günther, J., Mühlbauer, K., Seyfert, H. M., Schuberth, H. J., Hussen, J., Sauter-Louis, C., Hafner-Marx, A., & Zerbe, H. (2016). Early transcriptional events in the udder and teat after intra-mammary Escherichia coli and Staphylococcus aureus challenge. Innate Immunity, 22(4), 294–304. https://doi.org/10.1177/1753425916640057
- Philipsson, J., Ral, G., & Berglund, B. (1995). Somatic cell counts as a selection criterion for mastitis resistance in dairy cattle. Livestock Production Science, 41(3), 195–200. https://doi.org/10.1016/0301-6226(94)00067-H
- Politis, I., Zhao, X., McBride, B. W., & Burton, J. H. (1992). Function of bovine mammary macrophages as antigen-presenting cells. Veterinary Immunology and Immunopathology, 30(4), 399–410. https://doi.org/10.1016/0165-2427(92)90108-3
- Porcherie, A., Cunha, P., Trotereau, A., Roussel, P., Gilbert, F. B., Rainard, P., & Germon, P. (2012). Repertoire of Escherichia coli agonists sensed by innate immunity receptors of the bovine udder and mammary epithelial cells. Veterinary Research, 43, 14. https://doi.org/10.1186/1297-9716-43-14
- Prin-Mathieu, C., Le Roux, Y., Faure, G. C., Laurent, F., Béné, M. C., & Moussaoui, F. (2002). Enzymatic activities of bovine peripheral blood leukocytes and milk polymorphonuclear neutrophils during intramammary inflammation caused by lipopolysaccharide. Clinical and Diagnostic Laboratory Immunology, 9(4), 812–817. https://doi.org/10.1128/CDLI.9.4.812-817.2002
- Rainard, P., Cunha, P., Bougarn, S., Fromageau, A., Rossignol, C., Gilbert, F. B., & Berthon, P. (2013). T Helper 17-Associated Cytokines Are Produced during Antigen-Specific Inflammation in the Mammary Gland. PLoS ONE, 8(5). https://doi.org/10.1371/journal.pone.0063471
- Rainard, P., Foucras, G., Boichard, D.,& Rupp, R. (2018). Invited review: Low milk somatic cell count and susceptibility to mastitis. Journal of Dairy Science, 101(8), 6703–6714. https://doi.org/10.3168/jds.2018-14593
- Rainard, P., Foucras, G., & Martins, R. P. (2022a). Adaptive Cell-Mediated Immunity in the Mammary Gland of Dairy Ruminants. In Frontiers in Veterinary Science, 9. Frontiers Media S.A. https://doi.org/10.3389/fvets.2022.854890
- Rainard, P., Fromageau, A., Cunha, P., Gilbert, F., & Gilbert, F. B. (2008). Staphylococcus aureus lipoteichoic acid triggers inflammation in the lactating bovine mammary gland. Veterinary Research, 39(5), 52. https://doi.org/10.1051/vetres:2008034¨ı
- Rainard, P., Gilbert, F. B., & Germon, P. (2022b). Immune defenses of the mammary gland epithelium of dairy ruminants. Frontiers in Immunology, 13. https://doi.org/10.3389/fimmu.2022.1031785
- Rinaldi, M., Li, R. W., Bannerman, D. D., Daniels, K. M., Evock-Clover, C., Silva, M. V. B., Paape, M. J., Van Ryssen, B., Burvenich, C., & Capuco, A. V. (2010). A sentinel function for teat tissues in dairy cows: Dominant innate immune response elements define early response to E. coli mastitis. Functional and Integrative Genomics, 10(1), 21–38. https://doi.org/10.1007/s10142-009-0133-z
- Riollet, C., Rainard, P., & Poutrel, B. (2001). Cell subpopulations and cytokine expression in cow milk in response to chronic Staphylococcus aureus infection. Journal of Dairy Science, 84(5), 1077–1084. https://doi.org/10.3168/jds.S0022-0302(01)74568-7
- Rupp, R., Beaudeau, F., & Boichard, D. (2000). Relationship between milk somatic-cell counts in the first lactation and clinical mastitis occurrence in the second lactation of French Holstein cows. Preventive Veterinary



Medicine, 46(2), 99–111. https://doi.org/10.1016/S0167-5877(00)00142-2

- Rupp, R., & Foucras, G. (2010). Genetics of Mastitis in Dairy Ruminants. In S. C. Bishop, R. F. E. Axford, F. W. Nicholas, & J. B. Owen (Eds.), Breeding for Disease Resistance in Farm Animals (3rd ed., pp. 183–212).
- Sarikaya, H., Schlamberger, G., Meyer, H. H. D., & Bruckmaier, R. M. (2006). Leukocyte populations and mRNA expression of inflammatory factors in quarter milk fractions at different somatic cell score levels in dairy cows. Journal of Dairy Science, 89(7), 2479–2486. https://doi.org/10.3168/jds.S0022-0302(06)72322-0
- Schalm, O. W., & Lasmanis, J. (1968). The leukocytes: origin and function in mastitis. Journal of the American Veterinary Medical Association, 153(12), 1688–1694.
- Schalm, O. W., Lasmanis, J., & Carroll, E. J. (1967). Experimental Streptococcus agalactiae mastitis in cattle: attempts to superimpose the organism in lactating glands harboring unrelated bacterial infections and in glands with experimentally induced sterile inflammation. American Journal of Veterinary Research, 28(124), 685–695.
- Schukken, Y. H., Wilson, D. J., Welcome, F., Garrison-Tikofsky, L., & Gonzalez, R. N. (2003). Monitoring udder health and milk quality using somatic cell counts. In Veterinary Research, 34(5), 579–596. https://doi.org/10.1051/vetres:2003028
- Sharma, N., Singh, N. K., & Bhadwal, M. S. (2011). Relationship of Somatic Cell Count and Mastitis: An Overview. Asian-Australian Journal of Animal Science, 24(3), 429–438.
- Shook, G. E., Kirk, R. L. B., Welcome, F. L., Schukken, Y. H., & Ruegg, P. L. (2017). Relationship between intramammary infection prevalence and somatic cell score in commercial dairy herds. Journal of Dairy Science, 100(12), 9691–9701. https://doi.org/10.3168/jds.2017-12810
- Singh, K., Davis, S. R., Dobson, J. M., Molenaar, A. J., Wheeler, T. T., Prosser, C. G., Farr, V. C., Oden, K., Swanson, K. M., Phyn, C. V. C., Hyndman, D. L., Wilson, T., Henderson, H. V., & Stelwagen, K. (2008). cDNA microarray analysis reveals that antioxidant and immune genes are upregulated during involution of the bovine mammary gland. Journal of Dairy Science, 91(6), 2236–2246. https://doi.org/10.3168/jds.2007- 0900
- Soltys, J., & Quinn, M. T. (1999). Selective Recruitment of T-Cell Subsets to the Udder during Staphylococcal and Streptococcal Mastitis: Analysis of Lymphocyte Subsets and Adhesion Molecule Expression. In Infection and Immunity, 67(12). https://journals.asm.org/journal/iai
- Steeneveld, W., Hogeveen, H., Barkema, H. W., Van Den Broek, J., & Huirne, R. B. M. (2008). The influence of cow factors on the incidence of clinical mastitis in dairy cows. Journal of Dairy Science, 91(4), 1391–1402. https://doi.org/10.3168/jds.2007-0705
- Sumon, S. M. M. R., Parvin, M. S., Ehsan, M. A., & Islam, M. T. (2020). Dynamics of somatic cell count and intramammary infection in lactating dairy cows. Journal of Advanced Veterinary and Animal Research, 7(2), 314–319. https://doi.org/10.5455/JAVAR.2020.G423
- Sun, X., Zhao, R., Wang, N., Zhang, J., Xiao, B., Huang, F., & Chen, A. (2023). Milk somatic cell count: From conventional microscope method to new biosensor-based method. Trends in Food Science & Technology, 135, 102–114. https://doi.org/https://doi.org/10.1016/j.tifs.2023.03.020
- Suriyasathaporn, W., Schukken, Y. H., Nielen, M., & Brand, A. (2000). Low somatic cell count: A risk factor for subsequent clinical mastitis in a dairy herd. Journal of Dairy Science, 83(6), 1248–1255. https://doi.org/10.3168/jds.S0022-0302(00)74991-5
- Tadich, N. A., Carey, A., Porter, R., Ridley, J., Green, M. J., & Green, L. E. (1998). Case control study of risk factors for toxic mastitis in 26 dairy herds. Veterinary Record, 143, 362–365. http://veterinaryrecord.bmj.com/
- Van den Borne, B. H. P., Vernooij, J. C. M., Lupindu, A. M., van Schaik, G., Frankena, K., Lam, T. J. G. M., & Nielen, M. (2011). Relationship between somatic cell count status and subsequent clinical mastitis in Dutch dairy cows. Preventive Veterinary Medicine, 102(4), 265–273. https://doi.org/10.1016/J.PREVETMED.2011.07.013



- Vangroenweghe, F., Lamote, I., & Burvenich, C. (2005). Physiology of the periparturient period and its relation to severity of clinical mastitis. Domestic Animal Endocrinology, 29(2), 283–293. https://doi.org/10.1016/J.DOMANIEND.2005.02.016
- Vlasova, A. N., & Saif, L. J. (2021). Bovine Immunology: Implications for Dairy Cattle. In Frontiers in Immunology, 12. https://doi.org/10.3389/fimmu.2021.643206
- Yang, W., Zerbe, H., Petzl, W., Brunner, R. M., Günther, J., Draing, C., von Aulock, S., Schuberth, H. J., & Seyfert, H. M. (2008). Bovine TLR2 and TLR4 properly transduce signals from Staphylococcus aureus and E. coli, but S. aureus fails to both activate  $NF-<sub>k</sub>B$  in mammary epithelial cells and to quickly induce TNF $\alpha$  and interleukin-8 (CXCL8) expression in the udder. Molecular Immunology, 45(5), 1385–1397. https://doi.org/10.1016/J.MOLIMM.2007.09.004

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